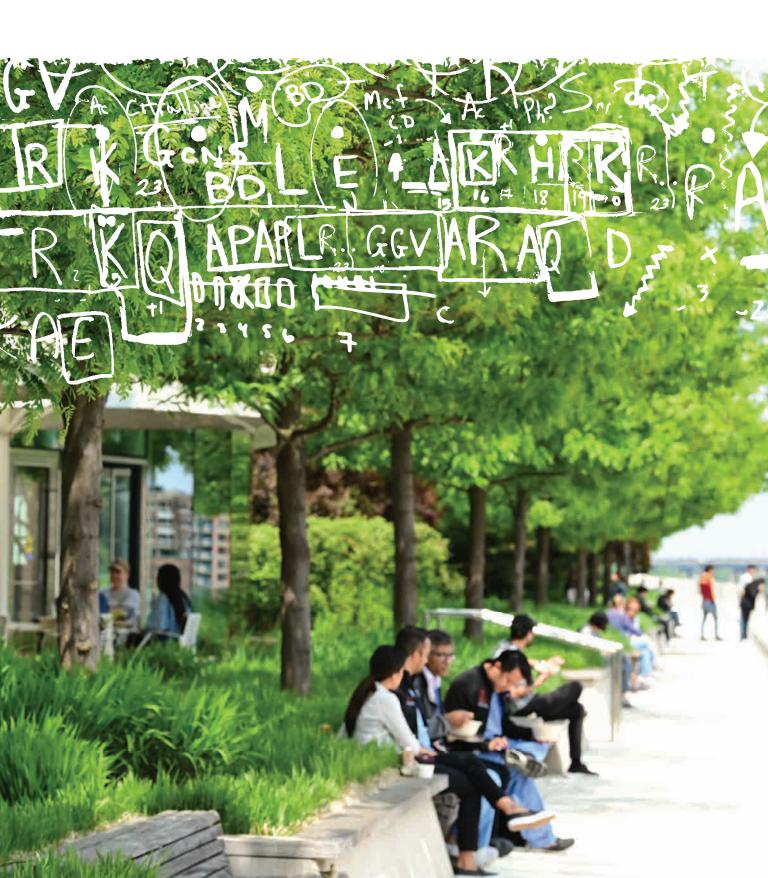
THE ROCKEFELLER UNIVERSITY

Schedule of Courses 2024–2025



The David Rockefeller Graduate Program offers a selection of courses, many of which students can choose based on their interests and area of thesis research. Organized by Rockefeller faculty and taught by scientists at the top of their fields, both from within and outside of the university, these courses provide a stimulating and dynamic curriculum that students can tailor to fit their personal goals in consultation with the dean of graduate studies.

Biochemical and Biophysical Methods, I & II

GREGORY M. ALUSHIN, SETH A. DARST, SHIXIN LIU, and MICHAEL P. ROUT

This course presents the fundamental principles of biochemistry and biophysics, with an emphasis on methodologies. In addition, case studies are discussed to examine how physical and chemical methods have been used to establish the molecular mechanisms of fundamental biological processes. The course is offered in two consecutive semesters. Part I introduces biological macromolecules and the experimental tools being used to dissect their three-dimensional structures and assembly principles. Part II covers methods aimed at delineating the conformational fluctuations, chemical turnovers, and kinetic trajectories of biological complexes at molecular, cellular, and evolutionary scales.

Class length and frequency: Two-hour session, twice weekly

Recommended reading: The Molecules of Life: Physical and Chemical Principles by John Kuriyan et al., Molecular Biology of the Cell by Bruce Alberts et al., Physical Biology of the Cell by Rob Phillips et al.

Method of evaluation: A five-minute oral presentation of a research proposal; a one-page written summary with specific aims for one semester and a three-page written review of a chosen method for the other semester

Bioinformatics

THOMAS CARROLL

In this course, the analysis of high-throughput sequencing using R and Bioconductor will be introduced. Students will learn the fundamentals of data handling in R, review the standard high-throughput sequencing data types, and manipulate this data using the Bioconductor R libraries. The course will then step through the processing and analysis of published RNA-seq, ChIP-seq, and ATAC-seq data.

Class length and frequency: Three-hour session, twice weekly Method of evaluation: Attendance and homework exercises twice weekly

Cell Biology

SANFORD M. SIMON and TIM STEARNS

This advanced course covering major topics in modern cell biology is taught by faculty and visitors who are specialists in various disciplines.

Class length and frequency: Three-hour lecture, once weekly; two-hour discussion, once weekly

Prerequisite(s): Good knowledge of textbook cell biology

Required reading: *Molecular Biology of the Cell* by Bruce Alberts et al., *Molecular Cell Biology* by James E. Darnell et al.

Recommended reading: Basic Histology by Luiz Carlos Junqueira et al.

Method of evaluation: Attendance, participation in the discussions, student presentations, and a final oral exam

Cell Biology of Nuclear Processes

AGATA SMOGORZEWSKA

The eukaryotic cell nucleus is a highly specialized organelle that carries the cell's genetic information and coordinates biological activities such as cell growth, metabolism, protein synthesis, and reproduction. This course will cover various aspects of nuclear function, with sessions focused on gene transcription (led by Robert G. Roeder), RNA processing and modification (Sohail Tavazoie), the nuclear envelope and nuclear pore complexes (Michael P. Rout), cell cycle control (Frederick R. Cross), DNA replication (Michael O'Donnell), DNA repair (Agata Smogorzewska), DNA damage response and telomeres (Titia de Lange), and chromosome segregation (Hironori Funabiki).

Class length and frequency: Two-hour lecture and discussion, once weekly

Prerequisite(s): Basic understanding of molecular biology and biochemistry

Required reading: To be assigned

Recommended reading: *Molecular Biology of the Cell* by Bruce Alberts et al. (selected chapters)

Method of evaluation: Attendance, participation in the discussions, and a take-home exam

Cell Cycle Control

FREDERICK R. CROSS and HIRONORI FUNABIKI

This seminar explores the current understanding of eukaryotic cell cycle control. Topics include the construction of a biochemical oscillator and overall structure of cell cycle control; positive and negative control of DNA replication; spindle morphogenesis and function; chromosome cohesion control; surveillance mechanisms (checkpoints) monitoring spindle and DNA integrity; and control of proliferation (start/restriction point control). The seminar relies heavily on studies in model organisms, but the emphasis throughout will be on aspects of cell cycle control conserved among eukaryotes.

Class length and frequency: 2.5-hour lecture and discussion, once weekly

Required reading: The Cell Cycle: Principles of Control by David O. Morgan; other readings as assigned

Method of evaluation: Attendance, homework exercises, and participation in the discussions

Cellular Neurophysiology

GABY MAIMON and VANESSA RUTA

This seven-week course will consist of 14 two-hour lectures with problem sets. The course will focus on the biophysical properties of neurons and electrically excitable membranes. Topics covered

will include the structure and function of ion channels, pumps/ transporters, and neurotransmitter receptors, and how these molecules contribute to resting potentials, action potentials, and synaptic transmission. The course will end with discussions on how biophysical principles can help to explain sensory and circuit-level properties in the nervous system.

Class length and frequency: Two-hour lecture and discussion session, twice weekly

Recommended reading: *Ion Channels of Excitable Membranes*, Third Edition, Bertil Hille; *Nerve, Muscle and Synapse*, Bernard Katz

Method of evaluation: Attendance, problem sets, and participation in the discussions

Cellular and Organismal Metabolism

KIVANÇ BIRSOY and PAUL COHEN

This course will cover the fundamental aspects of cellular and organismal metabolism, as well as exciting new applications for diseases such as obesity, diabetes, and cancer. Lectures will be given by the course directors and outside experts in the field.

Specific topics covered will include mitochondrial metabolism in the context of health and disease, lipids and non-polar metabolites in normal and disease physiology, signaling and metabolism, transcriptional regulation of metabolism, metabolic syndrome, exploiting metabolic pathways for cancer therapy, metabolomic approaches to studying cellular and organismal metabolism, and immunometabolism.

Class length and frequency: Two-hour lecture and discussion, once weekly

Prerequisite(s): Undergraduate biochemistry (recommended)

Required reading: Biochemistry textbook and discussion papers

Recommended reading: Lehninger Principles of Biochemistry by David L. Nelson and Michael M. Cox; Navigating Metabolism by Navdeep Chandel

Method of evaluation: Attendance, participation in the discussions, and presentations

Chemical Biology

TARUN KAPOOR

The spirit of this course is to explore the complexities of modern biology using the tools of chemistry. The lectures cover amino acid chemistry, nucleic acid chemistry, posttranslational modifications of proteins, discovery and use of chemical probes to examine cellular mechanisms, membrane chemistry, chemical tools for imaging, and natural product biosynthesis.

Class length and frequency: Two-hour lecture and discussion, once weekly

Recommended reading: Posttranslational Modification of Proteins: Expanding Nature's Inventory by Christopher Walsh; The Organic Chemistry of Biological Pathways by John McMurry and Tadhg Begley; Chemical Biology: From Small Molecules to Systems Biology and Drug Design, Volumes 1–3, by Stuart L. Schreiber et al.; Structure and Mechanism in Protein Science: A Guide to Enzyme Catalysis and Protein Folding by Alan Fersht; The Molecules of Life: Physical and Chemical Principles by John Kuriyan et al.

Method of evaluation: Attendance, participation in the discussions, and midterm and final exams

Experiment and Theory in Modern Biology

SANFORD M. SIMON and TIM STEARNS

This course introduces first-year graduate students to the methods and principles of current biological research. Students discuss preselected papers that illustrate methods of biological deduction and, with guidance from their faculty mentors, present and discuss papers and formulate conclusions about the experimental results. They also present a project based on one of the papers discussed in class. By the end of the course, students should be able to critically read a scientific manuscript and understand the principles used in interpreting scientific data.

Class length and frequency: Two-hour lecture and discussion, three times weekly in September

Required reading: To be assigned

Method of evaluation: Attendance, participation in the discussions, and student presentations

Fundamental Immunology and Microbiology

MING LI

This course provides a comprehensive overview of basic immunology beginning with innate immune responses, followed by a study of the main aspects of acquired immunity. Specific interactions of target cells and T cells that are regulated by the MHC molecule, and interactions between peptide antigens on the target cell and the antigen-specific T cell receptor, are studied. The generation and molecular structure of B and T cell antigen receptors, and signaling through immune receptors are covered in detail. Additionally, the development of antigen-specific T and B cells, and specific roles for some cytokines/lymphokines, are explored. The latter part of the course covers T and B cell-mediated immunity in more depth, along with topics of clinical relevance, such as microbial immunity, allergy, autoimmunity, tumor immunology, congenital and acquired immunodeficiencies, transplantation immunology, and immunotherapy. All these topics are studied though lectures and in-depth review of selected articles.

Class length and frequency: 3-hour session, twice weekly

Required reading: Janeway's Immunobiology, Ninth Edition, by Kenneth Murphy and Casey Weaver

Method of evaluation: Attendance, participation in the discussions, midterm and final exams

Fundamentals of Optics and Optical Microscopy for Biological and Biophysical Studies

ALIPASHA VAZIRI

This course provides a rigorous yet intuitive understanding of the physics behind optical microscopy, with a particular emphasis on applications in modern biological and biophysical research. Students will learn the basics of setting up and aligning simple optical systems and will build their own fluorescence microscope from individual components.

Overall, the course will introduce the fundamentals of geometric optics, including refraction, dispersion, Snell's law, lens maker equation, lens aberrations, matrix optics and ray tracing; the fundamentals of beam optics, including Gaussian beams, diffraction, and resolution; concepts from Fourier optics, including spatial filtering, 4f imaging, evanescent waves, and Abbe resolution limit;

microscopy, including light matter interaction, fluorescence and fluorescence microscopy, total internal reflection fluorescence (TIRF) microscopy, fluorescence confocal microscopy, two-photon microscopy, and calcium imaging; single-molecule fluorescence detection; and microscopy beyond the diffraction limit, including near-field imaging, stimulated emission depletion (STED) microscopy, and localization microscopy.

Class length and frequency: 1.5-hour lecture followed by a three-hour hands-on practical course, twice weekly over the course of three weeks

Prerequisite(s): A background in the natural sciences and basic knowledge of university-level calculus and physics

Method of evaluation: Class participation and laboratory performance

Genetics and Evolution

FREDERICK R. CROSS, DANIEL KRONAUER, and LI ZHAO

This seminar covers the basic mechanisms of genetics and evolution, including the generation of mutations and genetic segregation, linkage, and recombination (with an emphasis on linkage and segregation in eukaryotes). The course also considers changes in population genotypes when these basic genetic mechanisms are operating in the presence or absence of selective pressure. Changes in population genotypes can have effects ranging from polymorphism at neutral loci to the evolution of distinct species. Such changes are also used in historical analysis to trace migrations, evolution, and coevolution in diverse biological contexts.

Class length and frequency: 2.5-hour lecture and discussion, once weekly

Required reading: To be assigned

Method of evaluation: Attendance, homework exercises, and participation in the discussions

Introduction to Programming for the Life Sciences

SETH SYBERG

This course, which requires no previous programming knowledge, focuses on practical programming and data science skills using the Python programming language and associated scientific libraries. The course culminates in a final project directly applicable to the student's current research activities.

Class length and frequency: Two-hour lecture and lab, once weekly, and take-home programming assignments

Method of evaluation: Class participation, labs, assignments, and a final project

Mammalian Genetics

AGATA SMOGORZEWSKA

This course covers the genetics of bone marrow failure syndromes, cancer susceptibility, infectious diseases, obesity, diabetes, coronary heart disease, and neurodegenerative diseases. Also to be discussed are human gene mapping, disease modeling using mouse genetics, modern genetic tools including RNAi screening and genetic engineering using CRISPR, and ethical issues in modern human genetics.

Class length and frequency: Two-hour lecture and discussion, once weekly

Prerequisite(s): *Principles of Genetics*, Sixth Edition, by D. Peter Snustad and Michael J. Simmons (first six chapters)

Recommended reading: *Thompson & Thompson Genetics in Medicine*, Eighth Edition, by Robert L. Nussbaum et al.

Method of evaluation: Attendance, participation in the discussions, and take-home assignments

Mathematical Modeling

MARCELO O. MAGNASCO

This is an introduction to important topics in mathematical modeling and quantitative biology. A representative selection of subjects includes basic dynamics, fixed points and bifurcations, spiking neuron models, diffusion, chemical kinetics and systems biology, and stochastic simulation. Lectures introduce each topic and lab sessions cover programming methods in Python. The course is intended for students who have taken Math Review for Biologists or have a similar level of facility with calculus, linear algebra, and basic Python programming.

Class length and frequency: Two-hour lecture and discussion, once weekly; two-hour coding lab session, once weekly

Method of evaluation: Completion of a final project

Math Review for Biologists

MARCELO O. MAGNASCO

This is an intensive skill-development course, starting with calculus and linear algebra and leading up to differential equations, Fourier transforms, and related computational methods for model simulation. A concurrent journal club explores the major historical papers as well as contemporary biological modeling papers proposed by the students in full line-by-line detail.

Class length and frequency: Two-hour lecture and discussion, once weekly; two-hour journal club, once weekly

Method of evaluation: Attendance and participation in the discussions and journal clubs

Microbial Pathogenesis

LUCIANO MARRAFFINI and JEREMY M. ROCK

Infectious diseases continue to be a leading cause of human morbidity and mortality worldwide, and a major contributor to economic loss and poverty in developing countries. This course focuses on the molecular mechanisms of host-pathogen interactions and the pathogenesis of representative bacterial, fungal, and protozoan diseases. Topics include malaria, trypanosomiasis, toxoplasmosis, selected gram-negative and gram-positive bacterial infections, pathogenic mycobacteria, opportunistic mycoses, the evolution of pathogenicity, the impact of the host microbiota during microbial pathogenesis, and the development of antimicrobials and vaccines. The course is taught by Rockefeller and Weill Cornell Medicine faculty and selected guest speakers.

Each class includes a lecture followed by one or two student presentations on a paper suggested by the speaker in which the students outline follow-up experiments, and an optional lunch with the speaker.

Class length and frequency: Two-hour lecture and discussion, once weekly

Method of evaluation: Attendance, participation in the discussions, individual presentations, and a three-page research proposal on one of the topics covered in the lectures at the end of the course

Molecular Basis of Cancer

SOHAIL TAVAZOIE

This course is designed to teach concepts pertaining to the molecular regulation of cancer formation and progression. Each lecture will be accompanied by a review and a research article to be discussed over lunch.

Class length and frequency: Three-hour lecture and discussion, once weekly

Required reading: *The Biology of Cancer*, Second Edition, by Robert A. Weinberg; other materials as assigned

Method of evaluation: Attendance, participation in the discussions, and a take-home final exam

Quantitative Understanding in Biology Short Course

JASON BANFELDER and LUCE SKRABANEK

This course will prepare students to apply quantitative and statistical techniques to the analysis of experimental data. It emphasizes both practical and theoretical skills, and will involve hands-on exercises and homework using the GraphPad Prism program. Students will be well-positioned to meet the emerging requirements of funding agencies for formally planned experiments and fully reproducible and documented data analysis methods.

Specific topics include graphical, mathematical, and verbal communication of quantitative concepts; selection of appropriate statistical tests and interpretation of their results; design of appropriately sized experiments; formulation, evaluation, and analysis of mathematical models of biological function with an emphasis on linear and nonlinear regression; determination of model parameters; and critical comparison of alternative models with regard to overparameterization.

Class length and frequency: Nine two-hour sessions, once weekly

Required materials: Laptop with free GraphPad Prism software

Recommended reading: Intuitive Biostatistics by Harvey Motulsky; The Art of R Programming: A Tour of Statistical Software Design by Norman Matloff; R for Everyone: Advanced Analytics and Graphics by Jared P. Lander; Practical Computing for Biologists by Steven H.D. Haddock and Casey W. Dunn

Method of evaluation: Take-home problem sets

Responsible Conduct of Research

TRI-INSTITUTIONAL FACULTY and GUEST LECTURERS

Taught in collaboration with Memorial Sloan Kettering Cancer Center, this course raises awareness of ethical considerations relevant to the responsible conduct of research. It is aligned with requirements issued by the National Institutes of Health and the National Science Foundation. Attendance is mandatory for all first-year graduate students.

Class length and frequency: One-hour lecture and one-hour workshop, four times in the fall semester

Method of evaluation: Attendance, participation in the discussions, and online coursework

Science Diplomacy: The Context for Thinking Globally about the Biological and Medical Fields

JESSE H. AUSUBEL and MANDË HOLFORD

The shorthand term "science diplomacy" spans wide-ranging activities connecting science and technology with international affairs. In addition to global health and medicine, this course considers the larger context of dealing with nations in conflict, innovation in the public and private sectors, and views of science diplomacy from outside of the United States. This course will sample the current landscape of science diplomacy issues, programs, and organizations. Its goals are to help early-career biomedical scientists think more systematically about the global potential of their work including ethical, political, and economic implications; and to acquaint them with the people, networks, and resources available for scientific cooperation, including nations with whom cooperation may be especially challenging. Some students will be invited to join a field trip in March to Washington, DC, to meet with prominent science diplomacy practitioners and tour relevant institutions.

Class length and frequency: Two-hour lecture and discussion, once weekly for six weeks

Required reading: To be assigned

Method of evaluation: Attendance and participation in the

discussions

Seminars on Modern Biology

MEMBERS OF THE ROCKEFELLER UNIVERSITY FACULTY

This series of intensive seminars is designed to give incoming graduate students a chance to interact with Rockefeller faculty. Participation is mandatory for and limited to first-year students. In each session, two to four faculty members give brief introductions to their research and engage in a student-led discussion.

Social Evolution and Behavior

DANIFI KRONAUFR

This intensive course, held at The Rockefeller University Center for Field Research in Ethology and Ecology in Millbrook, New York, will include lectures, workshops, paper discussions, student presentations, and field outings. The course will explore complex questions from a variety of angles including genetics, behavioral ecology, ethology, neuroscience, and evolutionary and theoretical biology. It will cover several biological systems, ranging from single genetic elements, social microbes, insects, and vertebrates to mutualistic interactions between species.

Class length and frequency: One week of full-time participation

Required reading: To be assigned

Method of evaluation: Participation in the discussions, student

presentations, and hands-on workshops

Stem Cells in Tissue Morphogenesis and Cancer

ELAINE FUCHS and ALI H. BRIVANLOU

This course aims to present and discuss key concepts in stem cell biology, drawing on research from planaria to humans. It covers the basic principles of stem cells, from self-renewal and asymmetric cell divisions to tissue development, homeostasis, wound repair, and cancer. In addition to the basic lectures, there will be eight guest speakers who are world-renowned leaders in the field. Following each of these lectures, speakers will lead a discussion with the class on preassigned papers relating to the research topic.

Class length and frequency: Two-hour class lecture and discussion, once weekly; one-hour guest lecture, twice weekly

Required reading: To be assigned

Method of evaluation: Attendance, participation in the discussions, and a final written mini-review

Systems Biology: Process, Relation, and Scale

AMY E. SHYER and ALAN RODRIGUES

Systems biology has been defined as the study of biological systems whose behavior cannot be reduced to the linear sum of their parts. Today, systems biology has become synonymous with studies of large numbers of components. However, vast datasets are not sufficient for a systems approach. A systems biology approach also attempts to understand how the whole or system-level properties emerge from the interactions between the parts. Using this approach, this course will investigate the formation, maintenance, and breakdown of biological systems. Invited guests will participate in discussions grounded in specific research areas within the fields of developmental biology, regeneration, cell biology, cancer, and neuroscience.

Students are expected to absorb the fundamental principles of systems biology including self-organization, emergence, complexity, and causality in networks. They will also be expected to articulate how such principles could play a role in defining contemporary research questions in the lab.

Class length and frequency: Two-hour sessions, once weekly for

Required reading: To be assigned

Method of evaluation: Participation in weekly discussions, as well as a written paper

Systems Neuroscience

WINRICH FREIWALD

This 14-week course will consist of 14 two-hour lectures, with problem sets. The course will follow information processing from sensory to motor systems and focus on specific mechanisms and general principles. Topics will include olfactory, auditory, and visual information processing, memory systems, decision making, internal states, attention and executive functions, spatial navigation, and motor systems. The course will build on the cellular neurophysiology class. Basic knowledge of the biophysics of neurons and their compartments is expected.

Class length and frequency: Two-hour lecture and discussion session, once weekly

Method of evaluation: Attendance, problem sets, and participation in the discussions

Virology

CHARLES M. RICE, PAUL BIENIASZ, and THEODORA HATZIIOANNOU

In this course, Rockefeller faculty and guests give lectures and lead discussions about virology, with a major emphasis on the cellular and molecular biology of animal viruses. Topics include virus structure, replication, molecular genetics and gene expression, interactions with host cells, immunology, pathogenesis, viral vaccines, antiviral therapy, and resistance. A number of model systems are discussed, including cytocidal, steady state, and tumorigenic virus-cell interactions.

Class length and frequency: Two-hour lecture and two-hour journal club lunch discussion, once weekly

Required reading: To be assigned

Recommended reading: *Principles of Virology*, Volumes 1 and 2, Fourth Edition, by Jane Flint et al. (selected chapters); *Fields Virology*, Sixth Edition, by David M. Knipe and Peter M. Howley

Method of evaluation: Attendance, participation in the discussions, journal club presentations, and a written grant proposal

Supplementary Seminars, Tutorials, and Courses

Seminars and tutorials additional to those listed here are offered on demand. Students are welcome to speak with the dean of graduate studies to arrange training in any areas not covered by existing courses.

In addition, Rockefeller faculty and service departments offer a number of practical support courses. For example, students can learn about various aspects of computing and receive instruction in the efficient use of common software, as well as more complex computational tools.



SCIENCE FOR THE BENEFIT OF HUMANITY